

BEYOND A BETTER BOX™

XS-1000*i*[™] AUTOMATED HEMATOLOGY ANALYZER QUICK GUIDE



XS-1000*i* START-UP

Pre-Operation Checks

- 1. Refill printer with paper
- 2. Check waste container (if used)

Power on the Information Processing Unit (IPU)

- 1. Press the power button on the IPU.
- 2. For the Windows logon, "XS" is the user name and initially there is no password.* Press [ENTER] or click [OK].
- 3. When the XS IPU logon displays, input the user name and password, if required.
- 4. Press [ENTER] or click [OK].

NOTE: If a password is defined for the Windows log on, users have 3 chances to input the correct password. After 3 failed attempts, the account will be disabled for 30 minutes. Unlocking before 30 minutes requires administrative access (Call 1-888-879-7639).

Power on the Main Unit (MU)

NOTE: XS-Series Main Menu must be displayed before powering on Main Unit.

- 1. Power on Main Unit using the switch located on upper right side.
- 2. The XS performs self-checks. If any self-check fails, an error message displays. Refer to the Instructions for Use Manual chapter on Troubleshooting.
 - Microprocessor Check
 - Mechanical Parts Check
 - Temperature Checks
 - Background Check

Temperature Stability - XS-1000iCurrentTargetReaction Chamber37.640.7°cReagent Heater42.141.6°c



PU Logon			
User Name	ОК		
Password	Exit		



XS-1000*i* START-UP

NOTES ABOUT SELF CHECKS:

- The "Temperature Stability" dialog box displays the temperatures of the reaction chamber and reagent heater until they stabilize. If the box has not closed 30 minutes after start up, contact Technical Assistance.
- Pressure and vacuum readings are monitored by the XS.
 If a pressure or

Pressure	Temperature	Clo	c.e.
0.06 MPa 0.0000	Reaction Chamber	33.9	se.
-0.03 MPa -0.0011	Reagent Heater	42.4	
	FCM Sheath-Temp.	21.8	
	Environment	21.1	
Laser Current	HGB		
LD driver 2.0 mA	Convert	2177	
	Aspiration Sensor Convert	1996	
Sensors			
SNS1 SNS2 SNS3 SN	SA SNS5 SNS6 SNS7	SNS8 SNS9 SNS10	
SNS11 SNS12 SNS13 SNS	14 SNS15 SNS16 SNS17	SNS18 SNS19 SNS20	
SNS21 SNS22 SNS23 SNS	24 SHS25 SNS26 SNS27	SNS28 SNS29 SNS30	
SNS31 SNS32 SNS33 SNS	34 SNS35 SNS36 SNS37	SNS38 SNS39 SNS40	
SNS41 SNS42 SNS43 SNS	44 SNS45 SNS46 SNS47	SNS48	

vacuum error occurs, an error message displays. Click [Accept] on the Help dialog box to display the sensor dialog box. Follow the instructions.



- If the background check is successful, the Analyzer Icon at the bottom of the IPU screen turns green and the "Ready LED" on the Main Unit turns green.
- If any background count is greater than the acceptable limits, a "Background Error" displays and the Background Check dialog box opens. Parameters that exceed acceptable limits are displayed in red.
- If the "Background Error" displays, click [Accept] on the Help dialog box to perform another Autorinse.

Backgroun	d Check Results	Limit	Clos
квс Г	0.00	0.02 10/6/uL	
нав Г	0.0	0.1 g/dL	
PLT [0	10 10^3/uL	
NBC-C	0.02	0.30 10^3/uL	
WBC-D	0.00	0.1010^3/uL	

Troubleshooting

If any problems occur during Start-Up, QC Analysis, or Sample Processing, refer to Chapter 10, Troubleshooting in the Instructions for Use Manual.

XS-1000*i* QC ANALYSIS

QC Analysis (e-CHECK XS or e-CHECK XE)

- Warm controls to room temperature for at least 15 minutes.
- Mix controls by gentle, end-to-end inversion until re-suspended.
- DO NOT place on a mechanical mixer or rocker.
- "Control Entry Error" occurs if lot is not registered.

Manual Mode (4.5mL or 1.5mL vials)

- 1. From the Main Menu and with "Ready LED" green, click the [Manual] Icon or the [F2] function key on the keyboard.
- 2. Enter the QC lot number into the "Sample No." field using one of the following:
 - Ue the handheld barcode reader to scan the barcode label (*e*-CHECK XE only).
 - OR, Use the keyboard to manually type the number.
 NOTE: The prefix "QC" must always be uppercase and foll
- Click [OK] or press [Enter] on the keyboard. The QC dialog box opens.
 NOTE: If lot number has not been established, "Control Entry ERR" will display in the Help dialog box. Click [Accept] and [Close]. Follow QC New Lot Registration directions.
- 4. Verify appropriate lot and level is displayed.
- Select the appropriate sample tube adapter, and place it in the sample position area. Align the red dots, then turn the adapter clockwise until there is a click (turn about 45°) to secure it.
- 6. Mix the QC vial through gentle, end-to-end inversion, then place it into the XS sample adapter.
- 7. Press the [Start] switch on the right side above the sample position cover.
- 8. The QC results will display in the dialog box when sample measurement has
 - completed. Out of range results will display with red background. Press [Accept] for results to plot or [Cancel] to abort.
- 9. Repeat steps 1-8 for the remaining levels of QC to be analyzed.











XS-1000*i* QC ANALYSIS

QC without Barcode Label

- From the Main Menu and with "Ready LED" green, click the [Manual] Icon or press [F2] on the keyboard.
- 2. Click [QC] menu button in the Manual Sample No. dialog box.
- 3. Highlight the Control level and lot number to be analyzed.
- 4. Click [OK]. The QC dialog box opens.
- 5. Verify appropriate lot and level is displayed.
- Select the appropriate sample tube adapter, and place it in the sample position area. Align the red dots, then turn the adapter clockwise until there is a click (turn about 45°) to secure it.
- 7. Mix the QC vial through gentle, end-to-end inversion, then place it into the XS sample adapter.
- 8. Press the [Start] switch on the right side above the sample position cover.
- The QC results will display in the dialog box when sample measurement has completed. Out of range results will display with red background. Press [Accept] for results to plot or [Cancel] to abort.
- 10. Repeat steps 1-9 for the remaining levels of QC to be analyzed.

Vials of *e*-CHECK XS and *e*-CHECK XE are shown below:



			Can	cel
Discrete C CBC	CBC+DIFF	Capillary Mode C Yes ⓒ No		ic.
Patient ID		Patient Name		

File No.	Material	Lot No.	Exp. Day	Last QC measurement	-	OK
QC05	Control Level1	QC-60860804	06/27/2006	06/07/2006 14:29:55		
QC06	Control Level2	QC-60860806	06/27/2006	06/07/2006 14:47:14		Cancel
QC07	Control Level3	QC-60860807	06/27/2006	05/31/2006 11:10:49	1	
QC08	Control Level1	QC-61370801	07/21/2006	06/13/2006 15:04:03		
QC09	Control Level2	QC-61370802	07/21/2006	06/13/2006 14:55:45		Manual
QC10	Control Level3	QC-61370803	07/21/2006	06/13/2006 14:56:55	-	
QC11	Other1	QC-123	06/30/2006			
QC12	Other2	QC-11111	06/30/2006	06/01/2006 13:29:29		
QC17	Control Level3	QC-61370803	07/21/2006			
QC18	Control Level1	QC-61370801	07/21/2006		-	
0C19	Control Level2	QC-61370802	07/21/2006		-	

L-J - XS-1000i				
Nickname XS-1 File No. QCO8 Material Cont Lot No. QC-6 Exp. Day 0772	0001 rol Levell 1370801 1/2006			Accept Cancel Graph
RBC	10^6/uL g/dL % fL pg g/dL 10^3/uL fL % fL % fL %	WBC-C WBC-D NEUT# LYMPH# BASO# NEUT% LYMPH% MON0% EO% BASO% FSC-× DTEF-×	10A3/uL 10A3/uL 10A3/uL 10A3/uL 10A3/uL 10A3/uL 10A3/uL % % % % % % % % % % % % % % % % % % %	

XS-1000*i* SAMPLE PROCESSING

Manual Mode

- 1. From the Main Menu and with "Ready LED" green, click the [Manual] icon, or press [F2] on the keyboard.
- 2. Enter the patient identification into the "Sample Number" field using:
 - Handheld barcode reader to scan the patient label OR
 - Use the keyboard to manually enter the number
- 3. Select discrete tests if the LIS is not providing order information.
- 4. Click [OK], or press [Enter] on the keyboard.
- 5. Select the appropriate sample tube adapter, and place it in the sample position area. Align the red dots, then turn the adapter clockwise until there is a click (turn about 45°) to secure it.
- 6. Mix the patient sample through gentle, end-to-end inversion, then place it into the XS sample adapter.
- 7. Press the [Start] switch on the right side above the sample position cover.

Micro Tube Analysis in Manual Mode

NOTE: Remove the cap on the micro tube before performing analysis.

 From the Main Menu and with "Ready LED" green, click the [Manual] icon, or press [F2] on the keyboard.



- 2. Enter the patient identification number into the "Sample Number" field.
- 3. Select the Discrete tests to be performed.
- 4. Click [OK] or press [Enter] on the keyboard.
- Select the appropriate sample tube adapter, and place it in the sample position area. Align the red dots, then turn the adapter clockwise until there is a click (turn about 45°) to secure it.
- 6. Mix the sample through gentle, end-to-end inversion.
- 7. Remove the sample cap, then place the sample into the XS sample adapter.
- 8. Press [Start] switch on the right side above the sample position cover.
- 9. Repeat these steps to analyze additional micro tubes.







XS-1000*i* SAMPLE EXPLORER

Finding a Sample

- 1. Click [Explorer], or press [F7] on keyboard.
- 2. Deselect [Last 20] to access all stored data samples.
- 3. Either press [Ctrl]+[F] or click [Edit(E)] and select [Find(F)]; the "Find" dialog box appears.
- 4. Enter the Sample Number or Patient ID (number entered must match search number exactly, including leading zeros) If a number or letter is unknown, use
 "?" in its place .
- Click [NEXT] to search below the highlighted sample, or [PREV] to search above that sample. The located sample will be backlit in blue.
- 6. Select [Close] to exit the "Find" dialog window.



Modify a Sample

- 1. Click [Explorer] or press [F7].
- 2. Deselect [Last 20] to access all stored data samples.
- 3. Click on a Sample ID number to edit.
- 4. Click [Validate] or [F11] to un-validate the sample. (The 'V' in the far left column disappears).
- 5. Either click [Edit(E)] or [F10], then [Modify(P)]; a dialog box displays with the selected Sample ID.
- 6. Modify the Sample ID information, then click [OK]; the edited sample information is saved.
- 7. Click [Validate] or [F11] to re-validate the sample. If applicable, results will automatically print and retransmit to the Host.

Reprint to Graphic Printer or Retransmit Data to Host

NOTE: System cannot reprint or retransmit data that has not been validated (V).

- 1. Click [Explorer] or press [F7]. Deselect [Last 20] to access all stored samples.
- 2. Select Sample ID number(s) to reprint or retransmit to the Host.
 - To select any single sample, click the Sample ID number.
 - To select multiple samples (not necessarily in order), press and hold [CTRL] on the keyboard and click each additional sample.
 - To highlight a block of samples, click on the first sample to output, then press and hold [Shift] and click on the last sample to output.
- 3. To reprint, either click [Report(P)] or press [F12], then select [Report(GP)].
- 4. To retransmit, either click [Report(P)] or press [F12], then select [Host(HC)].





Edit(E) View(V) Rec Select All(A) Ctrl+A

Sample Explorer Filter[ALL]

Ctrl+F

Select All(A) Ctrl+A

Find(F)



XS-1000*i* QC SETUP

New Lot Registration

Empty QC Files:

- 1. From the Main Menu, click [QC Files] or press [F5].
- 2. Select an empty file number (1-20).
- 3. Click [Input] or press [F9].
- 4. Select the appropriate Control Level from the [Material] drop-down box.
- 5. Manually enter the lot number.
- 6. Select the appropriate expiration date.
- 7. Enter the Limit Range % for each parameter.
- 8. Verify that the information has been input correctly.
- 9. Select all parameters, then click [Variable Target].
- 10. Click [OK]. Repeat these steps to register remaining levels of new control.

Restore QC Files:

- 1. From the Main Menu, click [QC Files] or press [F5].
- 2. Select an empty file number (1-20).
- 3. Click [Record(R)], then select 'Restore(R).'
- 4. Open the [EB Control Limits] desktop folder.
- 5. Select and open the desired target limit file; the limits, lot number, and expiration date are restored.
- 6. The QC Files screen returns. Ensure the correct file is highlighted, then click [Input] or press [F9].
- 7. Manually enter the lot number.
- 8. Select the appropriate expiration date.
- 9. Verify that the information has been input correctly.
- 10. Select all parameters, then click [Variable Target].
- 11. Click [OK]. Repeat these steps to register remaining levels of new control.

NOTE: Any of the three procedures above may be used to register new lots of QC.

Reuse QC Files:

- 1. From the Main Menu, click [QC Files] or press [F5].
- 2. Select a used file, then click [QC Chart] or press [F11].
- Click and drag the cursor to select all QC data, or press [Ctrl]+[A] to select all data points.
- 4. Click [Delete], then [OK].
- 5. Click [Input] or press [F9].
- 6. Enter the Lot Number from the QC vial or Assay Sheet.
- 7. Select the appropriate expiration date.
- 8. Verify the [Limit Range %] for each parameter.
- 9. Select all parameters, then click [Variable Target].
- 10. Click [OK]. Repeat these steps to register remaining levels of new control.

👩 Open					23
Look in:	🔒 EB Control L	imits	•	← 🗈 📸 ▼	
Ca .	Name	*		Date modified	Туре
Recent Places	[XS-1000i][[XS-1000i][[XS-1000i][QC-11111]_March QC-22222]_March	2014_LSS-90-5 2014_LSS-90-5	6/3/2016 10:17 AM 6/1/2016 10:07 AM	QCF File QCF File
Desktop	[X2-10001][QC-33333J_March	2014_LSS-90-5	6/3/2016 10:17 AM	QCF File
Libraries					
Computer					
Network					
	•	III			F
	File name:			•	Open
	Files of type:	QC Backup File	(*.qcf)	-	Cancel



XS-1000*i* QC SETUP

Establishing Target Values

Analyze each control level once and compare to assay sheet. Comparison is required for the first point only as instrument compares subsequent results to ongoing mean.

- Analyze New Lot control at least 10 times before Auto Setting Targets. For statistical integrity, the accumulation of points should occur over a number of days. Minimum of 5 days, 2 points per day.
- The results of the New Lot number may be compared to the Current Lot number by selecting [F11] and then selecting the appropriate file.

Auto Setting Target Values

- 1. From the Main Menu, click [QC Files] or press [F5].
- Select an appropriate file number (1-20) and click [QC Chart] or press [F11]; the QC Chart displays.
- Select the range of data to include in calculations by clicking and dragging on the green cursor; to select all data, press [Ctrl]+[A].
- 4. Observe that the Mean, SD and CV are displayed in right data column.
- 5. Click [Input] or press [F9].
- 6. Highlight all parameters.
- Click [Auto Setting]. Ensure that only [Target] is selected and that [Limit] is deselected.
- 8. Click [OK]; target values are now displayed.
- Verify that the values returned as your analyzer's target values fall within range for each parameter on the Assay Sheet.
- 10. Click [OK] to close and save.





XS-1000*i* reviewing qc

Viewing Radar Charts

- 1. On the IPU, click the [QC Files] icon or press [F5].
- 2. Click the appropriate file number (1-20).
- 3. Once the file is selected, the Radar Chart will display on the right side of screen.
- 4. Control Data is displayed in blue on Radar Chart.
- 5. Date and time of analysis are displayed to the left in the "Analysis Date" column.
- 6. Parameters that exceed the acceptable limit will have a red "X" displayed on the radar chart. The parameter name will be backlit in red and an error message will be displayed to the left of the file number.

Viewing Levey-Jennings (L-J) Control Charts

- 1. On the IPU, click the [QC Files] icon or press [F5].
- 2. Double click on the appropriate file (1-20). The L-J Chart will open when [QC Chart] icon is selected.
- 3. Parameters that exceed an acceptable limit will have a red "X" displayed as the plotted point. The parameter name as well as the result value will be backlit in red.
- 4. To view remaining parameters use the scroll bar on right side of screen or the down arrow key on keyboard.



Shift ALL	NUMBA Default Column Default Default <thdefault< th=""> <thdefault< th=""> <thdefa< th=""><th></th><th></th></thdefa<></thdefault<></thdefault<>		
_			n+10
Item	n 1990 11 1977 11 1977 11 1977 11 1977	Data	3D Rean CV
RDC		2.24	0.015
H08		6.0	0.38
ICT		18.1	0.07
64		50.8	0.51
icH.		26.8	1.60
C102		39,1	2.17
PLT		51	2.1





XS-1000*i* QUALITY CONTROL

Print QC Reports

- 1. From the Main Menu, click [QC Files] or press [F5].
- 2. Select a file number (1-20), then click [QC Chart] or press [F11]; the QC Chart displays.
- 3. Click and drag the dark green cursor to select the range of QC data to print.
- 4. Click [Report(P)] or press [F12] on the keyboard.
- 5. Select [Report(GP)] to print graphs and L-J charts, or select [Ledger(LP)] to print raw data ledgers.

Delete QC File

NOTE: Deleting the QC file erases the control level, lot information, and historical limit %'s.

- 1. Click [QC Files] from the Main Menu or press [F5].
- Select the appropriate file number (1-20).
 Do not open the file.
- 3. Click [Delete] icon. Dialog box displays: "1 file(s) will be deleted. Are you sure?"
- 4. Click [OK] to delete or [Cancel] to abort.

Backup QC Files

- 1. From the Main Menu, click [QC Files] or press [F5].
- 2. Select the file number (1-20) to back up. Do not open the file.
- 3. Click [Record(R)] from the menu bar, and select [Backup(B)].
- 4. Select the folder or disk where you want to store the file.
- 5. Click [SAVE]; the file name is generated automatically:
 - File name format: (XS-1000i^Serial Number)(Version)(Date-Time)(Lot No).qcf
 - File name example: (XS-1000i^11597)(00-15)(03152011-112212)(QC-10050812).qcf

) IPU - [U - [QC Files]						
File(F)	Edit(E) View(V)	Record(R) A	ction(A) Report(P) Sett	ing(S) Window(W)	Help(H) Ver	:00-06 User Name:syst	mex
1 Help	F2 F3 100	R Menu	QC Files Work list	F7 F8 F8 Explorer Browser	F9	QC Chart In	sight Upper Low
	Nickname	No.	Material	Lot No.	Regist. Date	Analysis Dat	te Exp. Day
	XS-1000i	QC01	Control Level1	QC-60810801	05/02/2006	05/18/2006 10:5	50:53 05/26/2006
	XS-1000i	QC02	Control Level2	QC-60810802	05/02/2006	05/30/2006 16:	57:20 05/26/2006
	XS-1000i	QC03	Control Level3	QC-60810803	05/02/2006	05/26/2006 09:	19:28 05/26/2006
	XS-1000i		Contract i sould	00 00300004	05 /05 /000/		04/16/2006
	×S-1000i	Delete				14:2	29:55 06/27/2006
	X5-1000 i	1				14:4	47:14 06/27/2006
	×5-1000i	11	tile(s) will i	11:1	10:49 06/27/2006		
	XS-10001	ALG	e you surer		Can	cel 15:0	04:03 07/21/2006
	×5-1000i					14:	55:45 07/21/2006
	×5-1000i	QC.10	CONCIDE LEVELS	40 02310003	03/31/2000	00/10/1000 16:1	12:34 07/21/2006
	X5-1000i	QC11	Other1	QC-123	05/31/2006		06/30/2006
	XS-1000i	QC12	Other2	QC-11111	06/01/2006	06/01/2006 13:	29:29 06/30/2006
	×S-1000i	QC13					
	×S-1000i	QC14					
	XS-10001	QC15					
	×5-1000i	QC16					
	XS-1000i	QC17	Control Level3	QC-61370803	06/05/2006		07/21/2006
	×5-1000i	QC18	Control Level1	QC-61370801	06/05/2006		07/21/2006
	×5-1000i	QC19	Control Level2	QC-61370802	06/05/2006		07/21/2006
	ME KARAJ	0.02.0					



XS-1000*i Insight*™

Save QAP Data Using Sysmex Insight

- Sysmex *Insight* icon MUST be used for saving QC data for QAP.
- Review data and if desired, edit (delete)
- control data prior to submission. Ensure all QC data is plotted and no analysis error data is included (- - -) or (+ + +).
- 1. Click [QC Files] icon on analyzer IPU.
- 2. Click to highlight file to be saved.
- 3. Click on *Insight* icon on Menu Bar.
- 4. "Save As" dialog box opens. Insert USB Flash Drive into USB port.
- 5. If a "Please insert into Drive A" message displays, click [Cancel].
- Verify that the USB Flash drive name appears in "Save in" box. If not, click [▼] arrow and select drive (Removable disk) from list. Click [Save].
- 7. Repeat steps 2 4 for other files to be saved.
- With USB Flash drive still inserted, verify QC data was downloaded. Verify lot numbers and extensions ending with "ins".
- 9. Remove USB Flash drive.

Submit QAP Data To Sysmex

- 1. Go to www.sysmex.com/crc. Click [Enter Insight].
- 2. Log on:
 - User name: Enter your *Insight* customer email address.
 - Password: Enter your *Insight* customer password. Click [Login].
- 3. Click [Submit QC Data].
- 4. Click [] arrow next to "Please select your Analyzer" and select analyzer from list.
- To "Select QC data file" location, click [Browse]. Click [] arrow next to [Look in] box. Select *Insight* data folder on Desktop or Flash Drive.
- 6. Click on one QC data file. Click [Open]. File name is displayed. Click [Submit Data File].
- 7. Repeat steps 5 6 for other QC files.
- 8. Click [View QC Data Report] to view lot-to-date report.
- 9. Click [Log off].



F9

Input

Regist. Date

2008/11/26

2008/11/26

2008/11/26

2008/12/05

2008/12/05

2008/12/05

2008/12/09

2008/12/18

Ver.:00-15 Liser Name:sysmex

F11 ()

QC Chart

Analysis Date

010/08/17 09:21:19

2010/08/17 09:22:24

2010/08/17 09:23:34

2010/08/13 17:26:04

2010/08/13 17:28:21

2010/08/13 17:30:39

2010/07/23 10:27:13

2008/12/18 2010/07/23 10:33:13 2010/08/08

Insight

2010/07/23 10:30:02 2010/08/08

Low

Exp. Day

2010/09/13

2010/09/13

2010/09/13

2010/11/08

2010/11/08

2010/11/08

2010/08/08

File(E) Edit(E) View(V) Record(R) Action(A) Report(P) Setting(S) Window(W) Help(H)

QC Files

F5 💜 🖓

Material

rol Level:

Control Level2

Control Level3

Control Level1

Control Level2

Control Level3

Control Level1

Control Level2

Control Level3

Work list

Explorer

Brov

Lot No.

-0175080

0C-01750802

OC-01750803

QC-02310801

0C-02310802

QC-02310803

QC-01370804

QC-01370805

QC-01370806

F4

Menu

No.

0C02

QC03

QC04

QC05

QC06

0C07

0008

0C09

Nickname

-1000i

XS-1000iC

XS-1000iC

XS-1000iC

XS-1000iC

XS-1000iC

XS-1000iC

XS-1000iC

XS-1000iC

Save As				2
Save jr: My Recent Documents Desktop My Documents	My Recent D My Recent D Desktop My Docum My Docum Cal Di DVD/CU Fernovs Shared [My Decu My	Disk (E:) occuments ents ter sk (C:) -RW Drive (D:) be Disk (E:) Documents merts k (Places IC ies	•∰ * <u>6</u> \$	
My Computer				
My Network Places	File <u>n</u> ame:	QC-01750801	_	<u>S</u> ave
	Save as type:	Sysmex Insight Files (*.ins)	-	Cancel





QC Data

- Submit QC Data
- Review Your QC Data

Report Center

- Group Comparisons
- Customer QC Reports

XS-1000*i* SCATTERGRAMS



DIFF Channel





DIFF Scattergram



Side Scatter

XS-1000*i* results interpretation

The XS-1000*i* is a screening device which judges and marks each sample as either POSITIVE or NEGATIVE. This judgment is based on the presence or absence of IP messages (flags). The system evaluates numerical data, scattergrams, and particle size distributions for 21 reportable parameters and generates flags when abnormalities are or may be present.

NOTE: If an abnormality exists, determine a corrective action and verify results according to your laboratory's protocol. Some corrective actions may include checking the specimen for clots, remixing and reanalyzing the sample, performing a smear review, reviewing the patient history, etc.





NEGATIVE (green backlit): Absence of analysis errors or IP messages with the sample.

The following indicators may appear after the data:

NOTE: Indicators are listed in priority order.

- @ Data exceeds the linearity range
- * Data of low reliability (the value may have been influenced); verify results following laboratory SOP
- +,- Value outside of the upper or lower limit

The following indicators may appear in lieu of data:

- "----" Analysis impossible. Data won't appear due to analysis error or abnormal data.
- "++++" Data exceeds display limit.
- " A blank space indicates there is no order.

POSITIVE (red backlit): Presence of numerical analysis errors and/or IP messages with the sample.

Parameter:	Linearity (Whole Blood)
WBC	0-400 x 10³/µL
RBC	0-8 x 10 ⁶ /µL
HGB	0.0 - 25.0 g/dL
НСТ	0.0 - 60.0%
PLT	0 - 5000 x 10³/µL

Action Message



Count DIFF-CH. Add a DIFF and repeat the analysis.

XS-1000*i* REAGENT REPLACEMENT

Help Message:

Replace Container CELLPACK (EPK) Replace Container STROMATOLYSER-4DL (FFD) STROMATOLYSER-4DL ™ Replace Container STROMATOLYSER-4DS (FFS) STROMATOLYSER-4DS ™ Replace Container SULFOLYSER (SLS)

Reag	gent:
------	-------

CELLPACK TM

SULFOLYSER TM

- xs-1000iC Error List

Action

Reagents Replacement is in progress.

0%

Replace the empty reagent container(s)

Open Expiration: Volume:

60 Days	10L / 20L
60 Days	5L
90 Days	42µL
60 Days / 90 Days	500mL / 5L

Cancel Reset Alarm

4 L

Analyzer Prompted Message

- When a reagent container is empty, an alarm sounds 1. and the "Help" dialog box is displayed on the IPU. Click [Reset Alarm] or press [F1] to silence the audible alarm. The "Help" dialog box will display the reagent that needs to be replaced.
- 2. Click [Execute] to open Reagent Replacement screen.
- 3. Once open, the reagent replacement screen will list the reagents used by the system. The reagent requiring replacement will have a "Replace" notification next to it.
- 4. Use the handheld barcode reader to scan the reagent barcode label. Updated information is displayed, i.e. lot number, expiration date and volume.

NOTE: If needed, manually enter reagent information on the right side:

- a. Enter the Lot # found on reagent label.
- b. Enter the "unopened" expiration date from reagent label.
- c. Enter the number of days in "Exp. After Opened" box.
- d. Refer to the package insert for product expiration once opened.
- Remove cap from new reagent container. 5. Using clean technique to avoid contamination, remove reagent tubing spout kit from empty reagent container and insert

a	nd pre	ss [0	K] key.	r(s),			
agents Replacement -) Reagent	<mark>(S-1000)</mark> Replace	Lat No	Exp. Date	Amount	Setting V Exchange		Execute
CELLPACK	Replace	C5384	08/05/2006	4 L	-Manual Input	C5384	Cancel
SULFOLYSER		C5002	07/02/2006	250 mL	Exp. Day	//	
STROMATOLYSER-4DL		C5002	07/09/2006	4.2 L	EXP. After Opened	60 Davs	
STROMATOLYSER-4DS		⊂5006	07/09/2006	30 mL			

Amount

					Setting	Execut
Keagent	кертасе	LOT NO	Exp. Date	Amount	🖬 Exchange	EACCOL
CELLPACK	Replace	C5384	08/12/2006	20 L	Lot C5384	Cance
SULFOLYSER		C5002	07/02/2006	250 mL	Exp. Day	
STROMATOLYSER-4DL		C5002	07/09/2006	4.2 L	EXP After Opened 60 Days	
STROMATOLYSER-4DS		c5006	08/12/2006	30 mL		
agents Replacer	nent is	; in pro	gress.		Amount 20 L	

into new reagent container. Initial and date new reagent container.

- 6. Click [Execute] to initiate priming.
- Once completed, the reagent information will be updated in the reagent log. 7.

XS-1000*i* reagent replacement

User Prompted Reagent Replacement

NOTE: This procedure may be used for troubleshooting reagent or result issues prior to an empty or expired reagent.

- 1. Click the [Controller] icon in the Main Menu, then click on [Maintenance].
- 2. Click on [Reagent Replacement] to open reagent replacement screen.
- 3. Once open, the reagent replacement screen will list the reagents used by the system. Select the reagent to be replaced.
- 4. Using handheld barcode reader, scan the reagent barcode label. Updated information is displayed, i.e. lot number, expiration date and volume.
- Confirm the Exchange box is checked and lot number information is correct.
 NOTE: If needed, manually enter reagent

information on the right side:

- a. Enter the Lot # found on reagent label.
- b. Enter the "unopened" expiration date from reagent label.
- c. Enter the number of days in "Exp. After Opened" box.
- d. Refer to the package insert for product expiration once opened.
- 6. Remove cap from new reagent container. Using

clean technique to avoid contamination, remove reagent tubing spout kit from empty reagent container and insert into

new reagent container. Initial and date new reagent container.

- 7. Click [Execute] to initiate priming.
- 8. Once completed, the reagent information will be updated in the reagent log and remaining reagent volume screen.

🕐 IPU	- [Reagent I	.og Term:/	All Reagent: Al	ij					
File(F) Edit(E) V	iew(V) Rec	ord(R) Action(A) Report(P) Setting(S) Windo	w(W) Help(H)	Ver.:00-06	User Name:sysmex		
F1	F2 MANUAL	F3 100	F4 F5 Menu Q	C Files Work list Explorer E	rowser Input		F12 Out	per Lo	WBT
No.	Date	Time	Logon User	Reagent	Lot No.	Exp. Date	Exp. Date after opened	Amounts	Entry Type
1	05/09/2006	16:37	X5	CELLPACK					Manual
2	05/09/2006	16:41	XS	SULFOLYSER					Manual
3	05/10/2006	12:39	admin	CELLPACK	C5384	10/05/2007	60Davs	20L	Manual
4	05/10/2006	12:42	admin	SULFOLYSER	C5002	07/10/2006			Manual
5	05/10/2006	12:49	admin	SULFOLYSER	C5002	07/02/2006	60Days	500mL	Manual
6	05/10/2006	12:49	admin	STROMATOLYSER-4DL	C5002	09/11/2006	60Days	5.0L	Manual
7	05/10/2006	12:49	admin	STROMATOLYSER-4D5	c5006	09/30/2006	60Days	42mL	Manual
8	05/18/2006	09:24	sysmex	SULFOLYSER	C5002	07/02/2006	60Days	500mL	Manual
9	05/18/2006	15:40	sysmex	CELLPACK	C5384	10/05/2007	60Days	20L	Manual
10	05/24/2006	16:32	admin	CELLPACK	C5384	10/05/2007	60Days	200	Manual
11	06/05/2006	15:54	admin	CELLPACK	C5384	10/05/2007	60Days	20L	Manual
12	06/06/2006	14:52	sysmex	CELLPACK	C5384		60Days	20L	Manual
13	06/13/2006	16:32	sysmex	CELLPACK	C5384		60Days		Manual
14	06/13/2006	16:39	sysmex	CELLPACK	C5384		60Days		Manual
15	06/13/2006	16:49	sysmex	CELLPACK	C5384		60Days	20L	Manual
16	06/13/2006	16:49	sysmex	STROMATOLYSER-4DS	c5006	09/30/2006	60Days		Manual

Reagents Replacement - >	(S-1000i				
Reagent	Replace	Lot No	Exp. Date	Amount	Setting Execute
CELLPACK	Replace			4 L	Lot C5384 Cancel
SULFOLYSER		C5002	07/02/2006	250 mL	Exp. Day / / 🔹 🖉
STROMATOLYSER-4DL		C5002	07/09/2006	4.2 L	EXP. After Opened 60 Days
STROMATOLYSER-4DS		⊂5006	07/09/2006	30 mL	
Reagents Replace	ment is	s in pro	gress.		Amount
		0%			



XS-1000*i* MAINTENANCE

Daily - Shutdown

NOTE: When Shutdown is performed, the detector and dilution lines are cleaned. Put the instrument through a shutdown cycle at the end of each day's analyses; once every 24 hours if the instrument is running continuously.

Shutdown process is in progress.



- From the Main Menu, double-click the [Shutdown] icon.
- Click [Execute]; the procedure takes approximately 2 minutes to complete.
- 3. After the Shutdown

procedure has completed, the "Shutdown" dialog box closes and the "Power Off" dialog box appears.4. Click the [Restart] icon to initiate an Autorinse, or follow the Weekly Power-off procedure.

The Shutdown process will take about 2 minutes.

70%

Weekly - Power-off

Power off the analyzer at least once per week.

- 1. Press the Power Switch on the Main Unit.
- 2. Click the red [X] to close the software.
- 3. Click the Windows (Start) icon, and select [Shutdown]. The analyzer completely powers off.
- 4. When ready, follow the steps for Start-up, on Page 2.

Monthly - Rinse Sequence

Perform monthly or every 1200 samples. This procedure cleans the optical detector block.

- 1. Click the [Controller] icon from the Main Menu.
- 2. Click [Maintenance] icon.
- 3. Click [Monthly Rinse]. A dialog box will appear.
- 4. Select the appropriate sample adapter, then set a tube containing a minimum 3mL of CELLCLEAN 5% Sodium Hypochlorite Solution into the sample position area. Align the red dots, then turn the adapter clockwise until there is a click (turn about 45°) to secure it.
- 5. Press the white manual [Start] switch to initiate cleaning.
- After the procedure completes (approximately 15 minutes), either Power-off the instrument or click [Restart].

Nonthly Rinse - XS-1000i	
Monthly Rinse process will take about 15 minutes.	Cancel
Please set CELLCLEAN(3ml) to the Tube Holder,	
then press Start switch.	
CAUTION!	
Do not use any detergent except CELLCLEAN.	
Monthly Rinse is in progress.	
0%	
[¹	





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